Biological studies of some nutritional formula on liver functions

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Abstract

The present study investigated the effects ofsome nutritional formula on liver functionsin rats . Twenty one male albino rats were divided into (3) groups (7) rats in each group. The first group was control which fed on basal diet only as a negative control. The other groups were received basal diet containing 15% from formula (1) which contained 60 % Whey protein , 20 % Carrots and 20% Rhubarb and formula (2) contained 60 % Soy Protein , 20 % Hibiscus and 20% Turmeric. Liver functions was assessed by estimation of plasma concentration of enzymes activities of aspartate amino transferase (AST), alanineamino transferase (ALT),lipid fraction (total cholesterol and triglyceride) and cholesterol fraction (HDL-c, LDL-c, VLDL-c). Results showed an improvement in case of tested formula (2) followed by formula (1) for the above parameters. So, this study concluded that liver functions in rats can be ameliorated by administration of 15% from formula (2) which contained 60 % Soy Protein , 20 % Hibiscus and 20% Turmeric.

Key words: Nutritional formula, liver functions - cholesterol fractions.

Introduction

Phytotherapy is the treatment and prevention of diseases using plants or plants part, such as leaves, flowers, roots, fruits, seeds, and rhizomes. Preparation made from them called medicinal plants, or herbs **(Weiss and Fintelmann, 2000)**. Many plants were suggested to ameliorate or care the liver diseases, among them were the birch, celandine, Dates palm, dates, rosemary, papaya, onion, Turmeric and lettuce **(Morsi, 1992)**. Medicinal plants have very important place as they not only maintain the health and vitality of human beings and animals, but also cure several disease, including liver disorders without causing any toxicity **(Govind and Madhuri, 2010)**.

Turmeric is one of the oxidation of human low-density lipoprotein (LDL)(Gulfraz et al.,2011).

The Rhubarb root contain a high, nicotinic acid (niacin) and vitamin A. The dietary fiber of 14 varieties of Rhubarb root has been shown to be as high as 6.4-11.5% depending on variety and degree of ripeness(Münzbergová, 2012).

Carrot (Daucuscarota L.) contained a several insults (Dias, 2012). The effect of aqueous suspension of Carrot (Daucuscarota L.)on carbon tetrachloride induced liver damage , CCL4 induced toxicity induced liver damage antagonize aqueous dose of 250-500 mg/kg suspension of Carrot (Daucuscarota L.) by raising the level of LDH (Lactate dehydrogenase) and lowering of Carrot (Daucuscarota L.)by raising the level of AST (aspartic transaminases) and ALT (L-alanine amino transfers) 5% seed of Carrot (Daucuscarota L.)given to albino mice to evaluate hepatro protective action against dimethyl lami-noaze-benzen induced liver carcinogenesis was studied and the results showed significant changes in the plasma level of alanine (AST) alkaline phosphate (ALP) , total protein and serum albumin which analyzed by malondialdehyde but there is no harmful effect of Carrot (Daucuscarota L.)on the liver moreover , it exerts hepatoprotective effect against hepatobiliary carcinogens because of their antioxidant property (Sun et al ., 2009). So, the present study was carried out to investigate biological effects of some nutritional formula contained Carrot (Daucuscarota L.), Turmeric purpurea, Rhubarb root with soy protein and whey protein on serum parameters of liver functions in rats.

Material and Methods

Materials

Plants

The tested plants were obtained from local market in Shebin El-kom, Menofia Governorate, Egypt. Whey protein and soy Protein were obtained from National Research Center in Cairo.

Chemical reagents

Reagent kits were purchased from Diamond Diagnostics (Egypt).

Experimental animals

Twenty one white male albino rats weighing about 180 ± 5g were used as experimental animals in the present investigation. They were obtained from the animal house of Research Institute of Ophthalmology, El-Giza, Egypt. They were kept under observation for one week (as adapted period) before the onset of the experiment. The animals were housed in stainless steel cages at normal atmospheric temperature (25 ± 5°C) and had a 12 h light-dark cycle. Food and water were consumed *ad libitum*.

Methods:

Preparation of plant powder

These plants were washed and dried in drying oven at 50 °C for 3 days, then crushed and milled as a dried powder.

Animals diet

The basal diet was prepared according to **AIN (1993)**. The vitamin mixture was prepared according to **Campbell (1963)**, while salt mixture was prepared according to **Hegsted *et al.* (1941)**.

Experimental design

Twenty one male albino rats (180 ± 5g) were randomly divided into 3 equal groups (seven rats each). All rats were fed on basal diet for one week before starting the experiment for acclimatization. After the adapted period, the initial weight was 205 ± 5g. Groups of rats were as the follows:

Group (1): Rats (n=7) were fed on basal diet only as control negative group.

Group (2) :Rats (n=7) were fed on formula 1 which contained 60 % Whey protein , 20 % Carrots and 20% Rhubarb .

Group (3): Rats (n=7) were fed on formula 2 which contained 60 % Soy Protein , 20 % Hibiscus and 20% Turmeric.

By the end of theexperimental periods (28 days),rats were scarified using diethyl ether anesthesia at fasting state . Part of the blood was taken to determine the level of serum glucose and other portion of blood samples was collected and allowed tocoagulate at room temperature;other portion of blood was added toit,EDTA (ethylenediaminetetraceticacid) and centrifuged at 3000r.p.mfor 15 minutes. Serum was carefullyaspirated and transferred into cleancovettubes and stored frozen at -20°Cuntilthe time of analysis.

Biochemical analysis:

Serum Alkaline phosphatase (ALP) was determined according tothe procedure of **(IFCC methods., 1983)**.Aspartate amino transferase (AST) or (GOT) glutamic -oxalo acetic trans aminase and glutamic pyruvic transaminase (GPT) or Alanine amino transferase (ALT) were carried out according to the method of **Henry(1974) and Yound (1975)**.Glucose was determined by enzymatic test according to **Tietz (1976)** and **Yound (1975)**. Enzymatic colorimetric determination of triglycerides was carried out according to **Fassati and Prencipe (1982)**. Total Cholesterol was determined by colorimetric method according to **Allain (1974)**.The determination of HDL was carried out according to

the method of **Fnedewaid (1972)** and **Gordonand Amer (1977)**. The determination of VLDL (very lowdensity lipoproteins) and LDL (low density lipoproteins) was carriedout according to the method of **Lee and Nieman (1996)** . Total immunoglobulin (IgG, IgM, IgE and IgA) determined by Radioimmunoassay as described by the method of **Patrono and Peskar (1987).**

Histopathological study: Livers of the scarified rats were dissected, removed, washed with normal saline and put in 10% formalin solution. The fixed specimens were then trimmed, washed and dehydrated in ascending grades of alcohol. The tissue specimens were cleared in xylene, embedded in paraffin, sectioned at 4-6 microns thickness, stained with Hematoxylen and Eosin (H and E) and then studied under an electronic microscope according to (**Carleton ,1979)**.

Statistical analysis

Statistical analysis were done using the Statistical Package for the Social Sciences (SPSS for WINDOWS, version 11.0; SPSS Inc, Chicago). Comparative analyses were conducted using the general linear models procedure (SPSS Inc). Values of $P<0.05$ were considered statistically significant.

RESULTS

1-Effect offeeding 15% tested formula on serum lipidsin normal rats.

Administration ofthe tested formula at 15% level caused significant decreases in serum of total cholesterol, triglycerides, LDL-c and VLDL-c compared to control group (Table 1). Serum HDL-c levels increased but not significantly by the administration of the formula 1 (G2). Rats that were given formula 2 (G3) showed significantly higher levels of HDL-ccompared to control group (G1). The value of other lipid parameters of formula 2 were higher than control group. The obtained results in the same line of **Crouse (1999)** who found that soy protein can decrease LDL-c, total cholesterol and increase the level of HDL-c. Also, **Gulfraz *et al. (2011)*** reported that tumeric reduce the absorption of lipids from diet .

Table (1): Effect offeeding 15% tested formula on serum lipids in normal rats.

Serum lipids	G1	G2	G3
Total cholesterol	75.43^{a} ±2.19	76.33^{a} ±3.15	75.47^{a} ±1.13
Triglycerides	76.48^{b} ±0.13	76.8^{b} 1.03±	81.4^{a} ±3.01
HDL-cholesterol	53.94^{a} ±0.12	47.87^{b} ±1.15	48.89^{b} ±0.04
LDL-cholesterol	20.2^{c} ±1.17	24.9^{a} ±4.34	22.5^{b} ±0.74
VLDL-cholesterol	1.29^{b} ±1.17	1.56^{a} ±4.34	1.28^{b} ±0.74

Values are mean±SD. Values in the same column sharing the same superscript letters are not statistically significantly different at ($p<0.05$)

2--**Effect offeeding 15% tested formula on serum lipids on liver functions in normal rats.**

From data presented in table (2) the administration of formula 2 (G3) significantly reduced AST and didn't effected on ALT level when compared with the other treatment groups. There is no significant differences between group 2 with 3. From the above results, it could be noticed that crude fiber in legumes is a group of indigestible carbohydrates. It can improve the function of the alimentary tract and also lower blood glucose ,cholesterol levels and liver functions (**Roberfroid , 2000)**.

Table (2): -Effect offeeding 15% tested formula on liver functions in normal rats.

Parameters	**G1**	**G2**	**G3**
AST(U/L)	27.8^{b} ±0.07	30.2^{a} ±1.11	32.5^{a} ±0.21
ALT(U/L)	19.8^{b} ±1.91	28.9^{a} ±1.41	27.4^{a} ±0.5

Values are mean±SD. Values in the same column sharing the same superscript letters are not statistically significantly different at ($p<0.05$)

3--Effect of feeding 15% tested formula on immunological productions in normal rats .

From table (3), it could be observed that administration of the formula 2 it is affect to rats activity (Group 3). The mixture of formula 2 induced significant increases in serum levels of immunological profile compared to control group. The other tested formula caused non significant changes in serum level of immunological productions.The main antioxidant compounds in formula 2 are vitamins C and E, phenolic compounds. So, different studies have shown that they have a protective antioxidant effect on immunity status, cancer and cardiovascular diseases**(Mallillin*et al.,* 2008 and Murty*et al.,* 2010)**

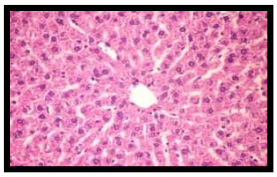
Table(3):Effect of feeding 15% tested formula on immunological productions in normal rats.

Immunological Profile mg/dl	G1	G2	**G3**
IgE	^{b} 59.87 ±1.34	^{b} 61.5 ±0.12	^{a} 65.17 ±0.25
IgM	^{b} 106.33 ±3.5	^{b} 108.65 ±0.65	^{a} 110.2 ±0.005
IgA	^{b} 106.5 ±1.5	^{b} 108.5 ±0.5	^{a} 113.1 ±0.1
IgG	^{c} 1089.66 ±11.16	^{b} 1095 ±19.87	^{a} 1101.05 ±8.05

Values are mean±SD. Values in the same column sharing the same superscript letters are not statistically significantly different at ($p<0.05$)

Histopathological examination of liver of the negative control rats fed on basal diet revealed normal histological picture of hepatic lobule which consists of central vein surrounded by normal hepatocytes as shown in (photo. 1). Examination of liver of group (2)showed of hepatocytes and infiltration of leucocytes in hepatic sinusoid (photo. 2). Liver and the third mixture showed marked improvements with no observed pathological lesions (photo 3). These results were according to **Mallillin*et al.*(2008) and Murty*et al.*(2010**) who found that tumericcan keep the liver tissue in normal status without any changes and improve the cells structure more than control group.

1



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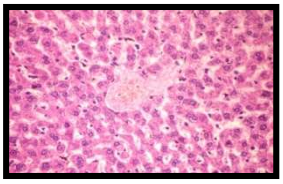

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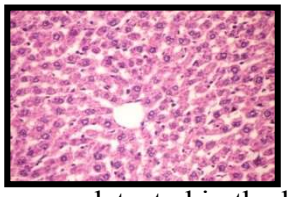

Photos (1): Histopathological changes detected in the liver of negative control, formula 1 and formula 2.

DISCUSSION

Several studies have showed that each of Soyprotien have long been recognized as an excellent source of high-quality protein. The soyprotien also contains a wide variety of chemical compounds that have potent bioactivity. Among these compounds are the isoflavones and the saponins. The goal of our research was to quantify isoflavone and saponin concentrations in elite soybean cultivars grown in different environments and to identify a naturally occurring high and low variety that could be used in animal studies of colon cancer. We observed significant environment × genotype interactions for the cultivars and selected 2 that provided the range of concentration for isoflavones and saponins. These were grown in an adequate quantity for animal studies, which are ongoing. They explored the influence of isoflavones and saponins on human colon tumor cells in culture, Caco-2, to determine

potential mechanisms through which these compounds influence the carcinogenic process. We observed the inhibition of Caco-2 cell proliferation by isoflavones and saponins, suggesting a protective effect of these compounds in colon cancer. Using purified soy saponins, we found no negative effects on mouse growth, organ weights, or intestinal morphology when the diet contained up to 3% saponins by weight. Hence, soy isoflavones and saponins are likely to be protective of colon cancer and to be well tolerated. Continuing studies will explore the cancer-protective effects of these compounds in animal models **(Ruth *et al* ., 2013).**

The insoluble carbohydrates in soyprotien consist of the complex polysaccharides cellulose, hemicellulose, and pectin. The majority of soybean carbohydrates can be classed as belonging to dietary fiber.

Within soybean oil or the lipid portion of the seed is contained thephytosterols: stigmasterol (17–21%), sitosterol(53–56%) and campesterol(20–23%) accounting for 2.5% of the lipid fraction.

Saponins, a class of natural surfactants (soaps), are sterols that are present naturally in a wide variety of food-plants: vegetables, legumes, and cereals–ranging from beans and spinach to tomatoes, potatoes and oats. Whole soybeans contain from 0.17 to 6.16% saponins, 0.35 to 2.3% in defatted soy flour and 0.06 to 1.9% in tofu. Legumes such as soybean and chickpeas are the major source of saponins in the human diet. Sources of non-dietary saponins include alfalfa, sunflower, herbs and barbasco. Recent studies have shown that saponins are potential functional food ingredients because of their physiological properties.

Soy contains isoflavones like genistein and daidzein. It also containsglycitein, an O-methylated isoflavone which accounts for 5–10% of the total isoflavones in soy food products. Glycitein is a phytoestrogen with weak estrogenic activity, comparable to that of the other soy isoflavones(**Teixeira et al. ,2000**).

Effect of tested formula on immunological its effect on increasing antioxydantenzyems could be indirect result of their effect on lipids metabolism.

The histopathological results showed that rats supplemented with formula 2 can prevent/reduce diet induce fatty liver. This fat reduction in the liver was confirmed by serum lipid analysis and by measurement of liver specific marker enzymes as mentioned before (**Teunissen *et al*.,2013).**

On the basis of the present results, it could be conclude that formula 2 which contained 60 % Soy Protein , 20 % Hibiscus and20% Turmeric especially at 15% may have synergistic effect and its intake of be useful for improving serum lipid profile, liver functions and immunological activity in rats. Moreover, this mixture has a promising effect on the liver tissues as it ameliorates the histopathological lesions seen in this organ of rats.

1- References

2- Ahmed, O.M.; Abdel Hamid, H.; Bastway, M. and Hasona, N.A. (2006):Antihyperglycemic effects of Cactus pear and Turmerics aqueous extract in diabetic and hypercholesterolemic rats. J. Egypt. Ger. Soc. Zool., 51A: 371-393.

3- AIN, (1993): American Institute of purified Diet for Laboratory Rodent, Final Report. J. Nutr., 123: 1939- 1951.

4- Allain, C. (1974): Cholesterol Enzymatic colorimetric Method. J. Clin. Chem., 20:470.

5- Ambrose, A. M.; DeEds, F. and Rather, L. J. (2009): Further studies on toxicity of CCl4٤ in rats. Proceedings of Society of Experimental and Biological Medicine, 74: 134-140.

6- An, J. H.;Seong,J.; Oh, H.; Kim, W.; Han, K. H. and Paik, Y. H. (2006): Protein expression profiles in a rat cirrhotic model induced byCCl٤. Korean J . Hepatol., 12: 93-102.

7- Ansari, R. A.; Tripathi, S. C.; Patnaik, G. K. and Dhawan, B. N. (1991): Antihepatotoxic properties of picroliv, an active fraction from rhizomes of Picrorhizakurroa. J. Ethnopharmacol., 34: 61-68.

8- Balasubramaniam, P.; Pari, L. and Menon, V. P. (1998): Protective effect of Turmeric (DaucusCarota L.) against lindane- induced hepatotoxicity in rats. Phytother. Res., 12(6): 434-436.

9- Bishayee, A.; Sarkar, A. and Chatterjee, M. (1995):Hepatoprotective activity of Turmeric (Daucuscarota L.) against carbon tetrachloride intoxication in mouse liver. J. Ethnopharmacology.; 47(2): 69-74.

10-Brandon, T. M. and David, M. B. (2012): Turmeric Bisacetylenicoxylipins-Phytochemicals behind the mask of the super food. J. Agric. Food chem., (10): 167-184.

11-Burtis, C.A. and Ashwood, E.R. (2001):Tietz's Fundamentals of Clinical Chemistry. WB Saunders, Philadelphia. 565.

12-Campbell, J. A. (1963): Methodology of protein evaluation R. A. G. Nutr. Document R. 10 Led. 37: June Meeting. New York.

13-Carleton, H. (1979): Histological Technique. 4 th Ed., London, Oxford University Press, New York, Toronto .

14-

15-Chieli, E. and Malvadi, G. (2008): Role of Cyt P-450 dependent and FAA containing mono oxygenases in the bioactivation of Thioacetamide and CCl٤, thiobezamide and their sulphoxides. Biochem.Pharmacol., 34: 395-396.

16-Crouse, J.R.; Morgan, T. and Terry, J.G. (1999) : A randomized trial comparing the effect of casein with that of soy protein containing varying amounts of isoflavones on plasma concentrations of lipids and lipoproteins. Arch. Intern. Med., 159:2070–2076.

17-Dias, J.S. (2012): Major Classes of Phytonutriceuticals in Vegetables and Health Benefits: A Review. Journal of Nutritional Therapeutics, 1: 31-62.

18-Fassati, P. and Prencipe, L. (1982): Triglyceride Enzymatic Colorimetric Method. J. Clin. Chem., 28:2077.

19-Fernandez - Lopez, J. A.; Almela, L.; Obon, J. M. and Castellar, R. (2010); Determination of antioxidant constituents in cactus pear fruits. Plant foods Hum. Nutr., 65(3): 253-259.

20-Fnedewaid, W.T. (1972): Determination of HDL. Clin. Chem., 8:499.

21-Fossati, P; Prencipe, L. and Berti, G. (1980): Use of 3.5- dichloro- z-hydroxybenzenesulfonic acid/4 aminophenazone chromogenic systems in direct enzymic assay of uric acid in serum and urine. Clin. Chem., 26:227-231.

22-Galati, E. M.; Mondello, M. R.; Giuffrida, D.; Dugo, G.; Miceli, N.; Pergolizzi, S. And Taviano, M. F. (2003): Chemical characterization and biological effects of Sicilian Opuntiaficusindica (L.) Mill. Fruit Juice: antioxidant and antiulcerogenic activity. J. Agric. Food Chem., 51(17): 4903-4908.

23-Galati, E. M.; Mondello, M. R.; Lauriano, E. R.; Taviano M. F.; Galluazzo, M. and Miceli, N. (2005):Opuntia ficus indica(L.) Mill. fruit juice protects liver from carbon tetrachloride- induced injury, phytother Res., 19(9); 796-800.

24-Gilani, A. H.; Janbaz, K. H. and Shah, B. H. (1998):Esculetin prevents liver damage induced by paracetamol and CCl4. Pharmacol Res., 37 (1):31-35.

25-Gordon, T. and Amer, M. (1977): Determination of HDL. J. Med., 62:707.

26-Govind, P. and Madhuri, S. (2010): Significance of Fruits And Vegetables In Malnutrition Cancer. PL Arch 10(2): 517-522.

27-Gulfraz, M. ; Sadiq, A; Tariq, H.; Imranim, M.; Qureshi, R. T. and Zeenat, A.(2011): Phytochemical analysis of seed of E. sativa. Pak. J. Bot., 43(2): 1351-1359.

28-Hegsted, D.; Mills, R. and Perkins, E. (1941): Salt Mixture. J. Biol. Chem., 138:459.

29-Henry, R. J. (1974): Clinical chemistry principles and techniques. 2ndED., Harper and publisher, New York, Philadelphia.

30-IFCC. (1983): Methods for measurement of catalytic concentration of Enzymes, parts 5: IFCC, methods for alkaline hosphatase. J. Clin. Chem. ClinBiochem., 21:731-748.

31-Ilavarasan, R.; Vasudevan, M.; Anbazhagan, S. and Venkataraman, S. (2003): Antioxidant activity ofthespesiapopulnea bark extracts against carbon tetrachloride-induced liver injury in rats. J. Ethnopharmacol., 87: 227-230.

32-Jayasekhar, P.; Mohanan, P.V. and Rahinam, K. (1997):Hepatoprotective activity of ethyl acetate extract of Acacia catechu. Indian. J. Pharmacology, 29:426-428.

33-Judzentiene, A. and Budiene, J. (2008): Volatile constituent from aerial part and roots of Cichoriumintybes L. (Chicory) grown in Lithuania. Chemija, 19(25): 25-28.

34-Kaur, N. and Gupta, A. K. (2002): Applications of inulin and oligofructose in health nutrition. Journal of Biosciences, 27 (7): 703 – 714.

35-Kowalska, D.P.; Feeley, R.M. and Jones, D.P. (1990): Use of exogenous glutathione for metabolism of peroxidized methyl linoleate in rat small intestine. J. Nutr., 120: 1115-1121.

36-Kruszynska, Y.T. and McIntyre, N. (1991): Carbohydrate metabolism. In: McIntyre, N.; Benhamou, P.J.; Bircher, J.; Rizzetto, M. and Rodes, J. (Eds.) Oxford Textbook of Clinical Hepatology. Oxford University Press, Oxford. 129-143.

37-Krylova, S. G.; Efimova, L. A.; Vymiatina, Z. K. and Zueve, E. P. (2006): The effect of cichorium root extract on the morphofunctional state of liver in rats with carbon tetrachloride induced hepatitis model. EKSP KlinFarmakol, 69(6): 34-36.

38-Kumar, C. H.; Ramesh, A.; Kumar, J. n. and Mohammed, I. B. (2011): A Review on hepatoprotective activity of medicinal plants, IJPSR, 2 (3): 501-515.
39-Lee, R. D. and Nieman, D. C. (1996): Nutritional Assessment. 2nd ED. Mosby, Missoun, USA.
40-Mansour, M. A. (2000): Protective effects of thymoquinone and deferrio-xamine against hepatotoxicity of carbon tetrachloride in mice. Life Sci, 66(26): 2583-2591.
41-MC Conn and Nakata, (2004): Glycoprotein (90kCa) isolated from Opuntiaficus-indicavar. saboten Makino lowers plasma lipid level through scavenging of intracellular radicals in triton WR-1339-induced mice. Biol Pharm Bukk., 29:1391-1396.
42-Morsi, A. E. (1992): Your Health and Healing between your Hands in Herbs. In Arabic, Egypt.
43-Muralidharan, P.; Balamurugan, G. and Kumar, P. (2008): Inotropic and Cardioprotective effects of Daucuscarota Linn.on isoproterenol- induced myocardial infarction. Bangladesh J. Pharmacol.,(3): 74-79.
44-Münzbergová, Z. (2012): Active constituents in Rheum acuminatum and Rheum australe (Polygonaceae) roots: A variation between cultivated and naturally growing plants. Biochemical Systematics and Ecology, 41:83-90.
45-Patel, S. (2013): Reviewing the prospects of Opuntia Pears as low cost functional Foods. Reviews in Environmental Science and Bio/Technology, 12(3):223-234.
46-Patrono, C. and Peskar, B. A.(1987): Radioimmunoassay in Basic and Clinical Pharmacology. Handbook of Experimental Pharmacology, Mishawaka, U.S.A.
47-Patton, C. and Crouch, S. (1977): Determination of serum urea Enzymatically Anal. Chem., 49.
48-Pieroni, A.; Janiak, V.; Durr, C. M.; Ludeke, S.; Trachsel, E. and Heinrich, M. (2002): In vitro antioxidant activity of non- cultivated vegetables of ethnic Albanians in southern Italy. Phytotheraphy Research, 16 (5): 467 – 473.
49-Potter, A. S.; Foroudi, S.; Stamatikos, A.; Patil, B. S. and Deyhim, F. (2011): Drinking Turmeric juice increases total antioxidant status and decreases lipid peroxidation in adults. Nutr.,10(1): 96.
50-Recknagel, R. O.; Glende, E. A.; Dolak, J. A. and Waller, R. L. (1989): Mechanism of Carbon-tertrachloride Toxicity". Pharmacology Therapeutics, 43 (43): 139-154.
51-Roberfroid, M. B. (2000): Prebiotics and legums: Are they functional foods? Am. J. Clin. Nutr., 71:1682S-1687S.
52-Rosen, H. R. and Keeffe, E. B. (1998): Laboratory evaluation of the patient with signs and symptoms of liver disease. In: Brandt LJ. (Ed.) Clinical Practice of Gastroenterology.Vol 2, Churchill Livingstone, Philadelphia, 812-20.
53-Ruth, S.; MacDonaldetall A.;Jimmy, D.;Browning, J.; George, E.; Rottinghaus, D. and Mark, A. (2013): Author Affiliations Environmental Influences on Isoflavones and Saponins in Soybeans and Their Role in Colon Cancer. The American Society for Nutritional Sciences.1: 123-129.
54-Salim, N.; Abdel Wahab, C.; Rabah, C. and Ahcene, B. (2009): Chemical Composition of Opuntiaficus- Indica (L.) fruit. African J. Biotechnology; 8(8): 1623-1624.

55-Schmidt, E.; Schmidt, F. W.; Mohr, J.; Otto, P.; Vido, I.; Wrogeman, K.; Herfarth, C. (1975): Liver Morphology and enzyme release. Further studies in the isolated perfused rat liver. In: Keppler (Ed.) Pathogenesis and Mechanism of Liver Cell Necrosis. Medical and Technical Publishing Co. Ltd., Lancaster. 147.

56-Sun, T., Simon, P.W. and Tamumuhardjo, S.A. (2009): Antioxidant Phytochemicals and Antioxidant Capacity of Biofortified Carrots (Daucus carota L.) of Various Colors. Journal of Agricultural and Food Chemistry, 57: 4142-4147.

57-Tapiero, H.; Tew, K. D.; Ba, G. N. and Mathe, G. (2002): Polyphenols: Do they play a role in the prevention of human pathologies? Biomed Pharmacother.,56: 200–207.

58-Teixeira , S.; Potter , S.M. and Weigel , R. (2000) : Effects of feeding 4 levels of soy protein for 3 and 6 weeks on blood lipids and apolipoproteins in moderately hypercholesterolemic men. Am. J. Clin. Nutr., 71:1077–1089.

59-Tesoriere, L.; Butera, D.; Pintaudi, A. M.; Allegra M. and Livrea M. A. (2004): Supplementation with cactus pear (Opuntiaficus-indica) fruit decreases oxidative stress in healthy humans: a comparative study with vitamin C. Am. J .Clin. Nutri.,80: 391-395 .

60-Teunissen, K.F. and Van Baak, M.A. (2013): The role of dietary protein in blood pressure regulation. Curr. Opin. Lipidol., 24:65-70.

61-Tietz, N.W. (1976): Fundamentals of Clinical Chemistry. Philadelphia, W.B. Saunders. P243.

62-Túnez, I.; Carmen Muñoz, M.; Villavicencio, M. A.; Medina, F. J.; de Prado, E. P.; Espejo, I.; Barcos, M.; Salcedo, M.; Feijóo, M. and Montilla, P.(2005):Hepato- and neurotoxicity induced by Thioacetamide and CCl4٤Protective effects of melatonin and dimethylsulfoxide. Pharmacol. Res., 52: 223-228.

63-Weiss, R. F. and Fintelmann, V. (2000): Herbal Medicine 2nd ED., Georg ThiemeVerlag, New York.

64-Yound, D. S. (1975): Determination of GOT. Clin.Chem., 21.

دراسات بيولوجية لبعض التوليفات الغذائية على وظائف الكبد

د/ حنان سعيد عبد السلام حجران

قسم التغذية وعلوم الأطعمة – مدرس مادة بكلية التربية النوعية جامعة المنوفية

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الملخص:

هذه الدراسة تستهدف معرفة تأثير بعض التوليفات الغذائية على وظائف الكبد في الفئران. وقد تم تقسيم واحد وعشرين من ذكور الفئران البيضاء إلى (٣) مجموعات وسبعه فئران (٧) في كل مجموعة. وكانت المجموعة الأولى المجموعة الكنترول التي تتغذى على الوجبة الضابطة فقط كمجموعة كنترول سالبة . والمجموعات الاخرى تتغذى على الوجبة الضابطة التي تحتوي على ١٥٪ من الوجبة الاساسية التوليفة التي تحتوى على ٦٠٪ بروتين شرش اللبن ، و ٢٠٪ من الجزر و٢٠% من الراوند والتوليفة (٢) تحتوي على ٦٠٪ بروتين الصويا، و ٢٠٪الخبيزة و 20٪. الكركم. تم تقييم وظائف الكبد عن طريق تقدير تركيز الإنزيمات من ترانسفيراز اسبارتاتي الأمينية (AST)، ألانين ترانسفيراز الأمينية (ALT)، الدهون (الكوليسترول والدهون الثلاثية)، الليبوبروتينات العالية والمنخفضة الكثافة . وأظهرت النتائج تحسنا نتيجة تناول التوليفة الغذائية رقم ٢ تليها التولفية الغذائية رقم ١ على القياسات السابقة. ولذلك هذه الدراسة اظهرت أن وظائف الكبد في الفئران يمكن تحسينها بواسطة ١٥٪ من التوليفة الغذائية ٢ والتى تحتوى على ٦٠٪ بروتين الصويا، و ٢٠٪ الخبيزة و 20٪ الكركم.

مفتاح الكلمات : التوليفة الغذائية، وظائف الكبد – مكونات الكوليستيرول.